baker's yeast reduction of anylalkyl and anylalkemil $\gamma-$ and $\delta-keto$ acids

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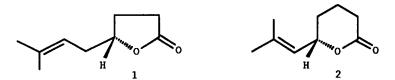
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Key Words: Baker's yeast; reduction; γ -lactones; δ -lactones.

<u>Abstract</u>: γ - and δ -Lactones 5, 6, 13, 14, 15 and 16 were synthesized via baker's yeast reduction of the corresponding keto acids 3,4 and 9-12. The enanticselectivity of the reduction is strongly dependent on the nature of the keto acid: the δ -lactones were always obtained in an eet higher than the γ -lactones and ranging from 70% to 100%.

A recent paper¹ on the synthesis of chiral γ - and δ -lactones 1 and 2 (Figure 1) from 7,7-dimethylbicyclo 3.2.0. hept-2-enone through enzyme-mediated procedures prompted us to report our approach to the same class of compounds. Our method is based on a straightforward baker's yeast mediated reduction of the corresponding γ - and δ -keto acids. The capacity of the microorganisms to enantioselectively reduce mid-chain aliphatic γ - and δ -keto acids is well known.^{2,3a} The majority of the microorganisms tested, including baker's yeast, afforded carbinols with (R) absolute configuration in high enantiomeric purity. Only a few examples yielding the (S) enantiomer are reported.

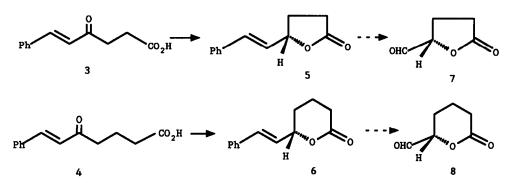
Figure 1



We first tested the ability of baker's yeast to reduce the carbonyl group of the unsaturated γ - and δ -keto acids 3 and 4, the aim being to obtain optically pure forms of

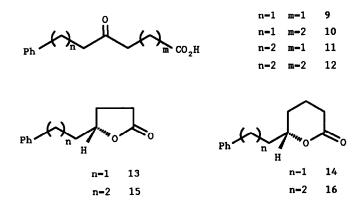
the stiryl lactones 5 and 6 (Scheme 1) of which the synthetic potential has recently been outlined.⁴ In fact, these lactones possess a versatile functionality, the double bond, which can be manipulated to provide synthetically useful structures such as the aldehydes 7 and 8. Although 7 can be prepared from L-glutamic acid⁵ the corresponding homologue 8 is not so readily accessible from chiral-pool components because its synthetic precursor, L-2-amino adipic acid, is a rare compound.

Scheme 1



The keto acids 3 and 4, prepared respectively from levulinic and 4-acetyl butyric acids by base catalyzed condensation with benzaldehyde, ⁶ were readily reduced by yeast. Unfortunately, the lactones 5 and 6 could be isolated in good chemical yield after acidification and extractive work up the stereochemical results of the reductions were rather disappointing, the lactone enantiomeric excesses being only 10% and 70% respectively. The optical purity was determined by ¹H NMR spectroscopy in the presence of Eu(hfc)₃ as the chiral shift reagent and by HPLC analysis on a chiral column.

Figure 2

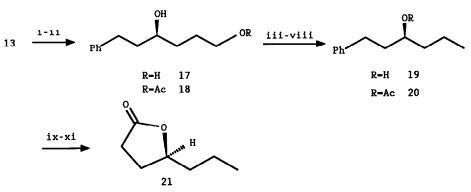


The absolute configuration (S) was established in both cases through a chemical correlation reported further on in the text. The modest enantioselectivity of the yeast

reduction of 3 and 4 is probably due to the presence of the conjugated double bond, in fact the yeast treatment of the keto acids 9 and 10^7 afforded the γ - and δ -lactones 13 and 14 in very high enantiomeric purity (Figure 2).

The absolute configurations and the optical purities were determined through chemical correlation. Thus the lactones 13 and 14 were transformed into known aliphatic lactones³ following the procedure reported for lactone 13 in Scheme 2. The correlation involved the transformation of the hydroxymethyl group of diol 17, obtained in turn via the LiAlH₄ reduction of the lactone 13, into a methyl group (e.g. 19). Initially, we tried to obtain this transformation via the reduction of the corresponding primary bromo derivative. Accordingly, the diol 17 was treated in methylene chloride at room temperature with NBS-Ph₃P.⁸ The reaction product, which can be isolated in 70% chemical derivative 22 with evidence of vield, was the tetrahydrofuran no other reaction-intermediates (Figure 3). The optical purity of derivative 22 was not determined, however the mechanism of this reaction will be further investigated. The desired conversion of 17 into 19 was achieved by the LiAlH4 reduction of the primary tosylate derivative obtained, in turn, through the protective group manipulations illustrated in Scheme 2. The alcohol 19 was acetylated and the aromatic ring of 20 was oxidized with ruthenium trichloride- H_5IO_6 , ⁹ yielding the (S)-heptanolide 21 in 92% ee. The absolute configuration and the optical purity of 21 were determined by glc analysis using a chiral capillary column, ¹⁰ and by comparison with an authentic sample. ^{3b} Thus, the absolute configuration of 13 is (R) and the enantioselectivity in the yeast reduction of γ -keto acid 9 is drammatically enhanced with respect to the unsaturated analog 3. The absolute configuration of 3 was established by catalytic hydrogenation of the double bond affording the (R) lactone 13 in 10% ee.

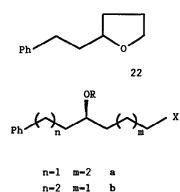
Scheme 2



i) $LiAlH_4$ -THF-reflux i1) $CH_2CHOCOCH_3$ -lipase-PhH-4h-RT iii)dihydropyrane-PTSA- CH_2Cl_2 iv) NaOH-MeOH-reflux-4h v) PTSCl-Py- CH_2Cl_2 vi) $LiAlH_4$ -THFreflux vii) HCl-MeOH-H₂O-reflux-3h viii) Ac₂O-Py-16h ix) RuCl₃H₂O-H₅IO₆- $CH_3CN-CCl_4-H_2O$ x) OH⁻ xi) H⁺

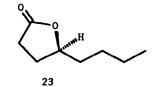
The δ -lactone 14 was transformed into the γ -octanolide 23 (Figure 3) as reported for 13 in Scheme 2. However, in this case, the treatment of the 1,5-diol 24 with NBS-Ph₃P did not afford the ring closure product, as reported for 17, but afforded, as expected, the primary bromo derivative. The tetrahydropyrane derivative can be obtained by treating the 5-hydroxy-1-bromo derivative with NaOH-MeOH.

Figure 3



с

n=2 m=2



X-OH R-H 24 a,b,c X-OAc R-H 25 a,b,c X-H R-H 26 a,b,c X-H R-Ac 27 a,b,c

Glc analysis on chiral column¹⁰ of γ -octanolide 23 showed (S) absolute configuration and 100% ee. Thus, the yeast reduction of γ -keto acid 10 afforded the lactone 14 with (R) absolute configuration, as for the reduction of 9, but in higher optical yield.

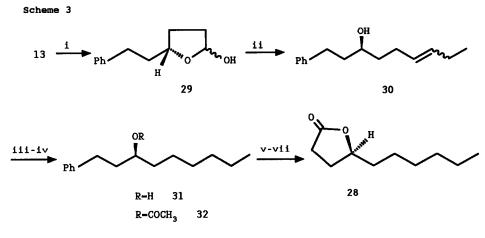
Catalytic hydrogenation of 6 led to the determination of its absolute configuration, affording 14 in (R) absolute configuration.

The lactone 13 was also correlated to the known lactone 28,¹¹ according to the sequence reported in Scheme 3, and this confirmed the (R) configuration of 13 (28: 97% ee by glc analysis).

The unsaturated lactone 5, obtained in low optical yield by yeast reduction, can be prepared by a simple manipulation of 13. In fact, lactone 13 after benzylic bromination (NBS, CCl₄) and dehydrobromination (DBU) afforded lactone 5 in the same optical purity as the starting material and in good chemical yield. On other hand, the transformation of δ -lactone 14 into the unsaturated δ -lactone 6 using this procedure did not perform so well, the chemical yield being very low (30%).

Subsequently, the γ - and δ -keto acids 11 and 12 were reduced by baker's yeast to give lactones 15 and 16 (Figure 2). These were submitted to the usual correlation protocol which afforded the aliphatic δ -lactones 33 and 34, respectively (Figure 4). The optical purities (40% for 33, and 93% for 34) and the absolute configuration ((S) in both cases) were determined by optical measurements and comparison with the $[\alpha]_D^{25}$ values reported in the literature^{3a} (33: $[\alpha]_D^{25}=-26.4^*$ (c 1, THF); 34: $[\alpha]_D^{25}=-58.8^*$ (c 1, THF)). Also in this case the δ -keto acid was reduced with higher enanticelectivity than

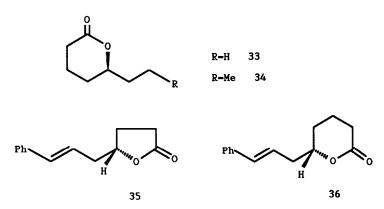
the γ one, as was observed for the reduction of 13 and 14.



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i) DIBAL ii) Ph_3PCHCH_2CH_3-THF iii) H_2-Pd-C-MeOH-H_2O iv) Ac_2O-
Et_3N-DMAP-CH_2Cl_2 v) RuCl_3H_2O-H_5IO_6-CH_3CN-CCl_4-H_2O vi) OH<sup>-</sup> vii) H<sup>+</sup>
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The unsaturated lactones 35 and 36 can be obtained in good yield from 15 and 16, respectively, by the benzylic bromination-dehydrobromination procedure.

Figure 4



These results confirm the validity of baker's yeast as a simple system for the production, through enantioselective carbonyl reduction, of quantities of materials with very high ee. At the same time, this further confirms that baker's yeast is very sensitive to structural limits, making the precise accetability of a substrate by the enzymic system(s) presiding over the carbonyl reduction unpredictable.

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EXPERIMENTAL

¹H NMR spectra were recorded with a Bruker AC-250 or a Bruker CXP-300 instrument in the FT mode or with a Varian EM-390 with tetramethylsilane as internal standard. Optical rotations were measured in a 1 dm cell of 1 ml capacity by using a Jasco DIP-181 polarimeter. Silica gel $60F_{254}$ plates (Merck) were used for analytical TLC, 270-400 mesh silica gel (Merck) for flash chromatography or 70-230 mesh silica gel (Merck) for chromatography. HPLC analyses were performed on a Merck Hitachi L-6200 with a Chiracel OD (Daicel) column and a UV (245 nm) detector using a Hewlett-Packard 3396A integrator. GLC analyses were performed on a Hewlett-Packard 5890 instrument with a capillary chiral Megadex 1 column using a Hewlwtt-Packard 5895A integrator. The keto acids 3-4 and 9-12 were synthesized according to literature procedures (see ref. 6 and 7).

Yeast Reduction.General Procedure. A suspension of 2.5 kg of baker's yeast and 2.0 kg of D-glucose in 7.5 l of tap water was stirred for 30 min at 32°C. A solution of keto acids (150 mmol) in 150 ml of 1M NaOH was then added. After 16h at room temperature 1.0 kg of Celite was added and the reaction mixture was filtered washing, the Celite pad with 2.0 l of AcOEt. The filtrate was adjusted to pH 4 with 2N HCl and extracted twice with 2 l portions of AcOEt. The organic layer was dried over sodium sulfate and the solvent was evaporated under reduced pressure. The crude material was chromatographed on 350 g of silica gel with n-hexane/AcOEt 8/2 as eluent, giving the lactones 5-6 and 13-16 in 70-80% yield.

<u>Lactone 5</u>: ¹H NMR (CDCl₃, 250MHz) δ : 2.00-2.20 (1H, m), 2.40-2.70 (3H, m), 5.06-5.20 (1H, m), 6.20 (1H, dd, J=15.00, 7.50Hz), 6.70 (1H, d, J=15.00Hz), 7.20-7.50 (5H, m). ee 10% (¹H NMR in the presence of Eu(hfc)₃ and HPLC with Chiracel OD column using n-hexane/iPrOH 9/1 as eluent flow 0.6 ml/min). Anal.Calcd. for C₁₂H₁₂O₂: C,76.57; H,6.44. Found: C,76.60; H,6.40.

Lactone 6: ¹H NMR (CDCl₃, 300MHz) δ : 1.70-1.82 (1H, m), 1.88-2.12 (3H, m), 2.46-2.70 (2H, m), 4.94-5.04 (1H, m), 6.19 (1H, dd, J=15.50, 6.00Hz), 6.65 (1H, d, J=15.50Hz), 7.20-7.40 (5H, m). ee 70% (determined as reported for 5). Anal.Calcd. for C₁₃H₁₄O₂: C,77.20; H,6.99. Found: C,77.22; H,6.98.

<u>Lactone 13</u>: ¹H NMR (CDCl₃, 250MHz) δ : 1.80-2.14 (3H, m), 2.24-2.38 (1H, m), 2.49-2.58 (2H, m), 2.64-2.90 (2H, m), 4.42-4.52 (1H, m), 7.10-7.34 (5H, m). [α] $_{D}^{25}$ =+71.3° (c 1.03, CHCl₃). Anal.Calcd. for C₁₂H₁₄O₂: C,75.76; H,7.43. Found: C,75.70; H,7.45.

<u>Lactone 14</u>: ¹H NMR (CDCl₃, 250MHZ) δ : 1.48-2.12 (6H, m), 2.38-2.42 (4H, m), 4.20-4.33 (1H, m), 7.10-7.34 (5H, m). $[\alpha]_D^{25}$ =+81.9* (c 1.04, CHCl₃). Anal.Calcd. for $C_{13}H_{16}O_2$: C,76.44; H,7.91. Found: C,76.47; H,7.95.

<u>Lactone 16</u>: ¹H NMR (CDCL₃, 250MHz) δ : 1.40-2.00 (8H, m), 2.33-2.70 (4H, m), 4.22-4.36 (1H, m), 7.12-7.34 (5H, m). $[\alpha]_D^{25}$ =+21.7° (c 1.04, CHCl₃). Anal.Calcd.for $C_{14}H_{18}O_{2}$: C,77.03; H,8.33. Found: C,77.00; H,8.37.

Diol 17. A solution of the lactone 13 (20 g, 105.3 mmol) in 30 ml of dry THF was added to a boiling THF (100 ml) suspension of LiAlH₄ (7 g, 184.4 mmol) under nitrogen. After refluxing for 4h, the reaction mixture was quenched by sequential addition of AcOEt, a saturated solution of potassium sodium tartrate and water. The organic layer was separated, dried over sodium sulfate and the solvent evaporated under reduced pressure. The crude product was purified by chromatography (AcOEt/n-hexame 65/35) to give diol 17 in 78% yield. ¹H NMR (CDCl₃/D₂O, 90MHz) δ : 1.50-1.95 (6H, m), 2.55-2.85 (2H, m),

3.50-3.78 (3H, m), 7.10-7.33 (5H, m). $[\alpha]_D^{25}$ =+7.5° (c 1.00, CHCl₃). Anal.Calcd. for $C_{12H_{18}O_{2}}$: C,74.19; H,9.36. Found: C,74.16; H,9.30.

<u>Acetyl derivative 18</u>. The diol 17 (14g, 72.2mmol) in 150 ml of benzene-n-hexane (1:1) was treated with vinyl acetate (13.4ml, 216.6mmol) and 3g of lipase from Candida cylindracea (Fluka) at room temperature. After 3h stirring, the reaction mixture was filtered and the solvent evaporated under reduced pressure. The crude product was purified by chromatography (n-hexane/AcOEt 8/2) to give 18 in 83% yield. ¹H NMR (CDCl₃, 250MHz) δ : 1.48-1.85 (7H, m), 2.03 (3H, s), 2.61-2.87 (2H, m), 3.60-3.70 (1H, m), 4.09 (2H, t, J=6.00Hz), 7.18-7.31 (5H, m). $[\alpha]_D^{2^5}=+6.0^*$ (c 1.00, CHCl₃). Anal.Calcd.for $C_{14}H_{20}O_{3}$: C,71.16; H,8.55. Found: C,71.12; H,8.58.

Alcohol 19. A solution of acetyl derivative 18 (14g, 59.3mmol) in methylene chloride (80 ml) was treated with dihydropyrane (5.9ml, 65.23mmol) and p-toluensulfonic acid (1g) at 0°C. After 3h, 3% NaHCO3 aqueous solution (20ml) was added and the organic phase was separated, dried over sodium sulfate and the solvent evaporated under reduced pressure. The crude product was dissolved in MeOH (50ml), treated with sodium hydroxide (4.0g, 100mmol) and refluxed for 4h. The reaction mixture was concentrated, diluted with ice-water and extracted with methylene chloride (2x100ml). The organic extracts were dried over sodium sulfate and the solvent evaporated under reduced pressure. A solution of the crude product was treated with p-toluensulfonyl chloride (19g, 100mmol) at 0°C. The reaction mixture was warmed at room temperature and, after 16h, was poured into ice-water then extracted with methylene chloride (3x100ml). The organic layers was washed with 3% HCl, water, 3% NaHCO3, water and dried over sodium sulfate. The solvent was evaporated under reduced pressure. The crude product was dissolved in THF (30ml) and added to LiAlH4 (5g, 131.6mmol) in boiling THF (100ml). After 10h, the reaction mixture was treated, in sequence, with AcOEt, methanol and potassium sodium tartrate. The solvant was evaporated under reduced pressure. The residue dissolved in 100 ml of MeOH/H $_{20}$ (1:1) was treated with conc. HCl (5ml) and refluxed for 3 h. The reaction mixture was concentrated, cooled and extracted with methylene chloride (2x100ml). The organic extract was dried and the solvent evaporated under reduced pressure. The crude product was purified by chromatography (n-hexane/AcOEt 9/1) to give the compound 19 in 39% yield. ¹H NMR (CDCl₃/D₂O, 250MHz) δ :0.91 (3H, t, J=5.50Hz), 1.28-1.53, (4H, m), 1.65-1.85 (2H, m), 2.60-2.87 (2H, m), 3.58-3.69 (1H, m), 7.15-7.32 (5H, m). [α]_D²⁵=+12.8° (c 1.00, CHCl₃). Anal.Calcd.for C12H180: C,80.85; H,10.20. Found: C,80.88; H,10.21.

<u>Acetyl derivative 20</u>. A solution of alcohol 19 (3g, 16.9mmol) in pyridine (4ml) was treated with Ac₂O (4ml, 42.4mmol) at room temperature. After 16 h, the reaction mixture was evaporated under reduced pressure and the crude product purified by distillation (b.p. 80°C/14mmHg) to give 20 in 90% yield. ¹H NMR (CDCl₃, 300MHz) &: 0.91 (3H, t, J=7.50Hz), 1.24-1.42 (2H, m), 1.49-1.53 (2H, m), 1.82.1.93 (2H, m), 2.03 (3H, s), 2.54-2.71 (2H, m), 4.90-5.00 (1H, m), 7.13-7.30 (5H, m). $[\alpha]_D^{25}=-16.5^{\circ}$ (c 1.00, CHCl₃). Anal.Calcd for C₁₄H₂O₂: C,76.33; H,9.17. Found: C,76.28; H,9.12.

Lactone 21. Compound 20 (2g, 9.09mmol) dissolved in acetonitrile (18.1ml) carbon tetrachloride (18.1ml) and water (30.3ml) was treated with periodic acid (29.2g, 129.0mmol). The reaction mixture was stirred 10 min (until both phases became clear) and then ruthenium trichloride hydrate (37.8mg, 0.18mmol) was added. After stirring for 3h, the reaction mixture was cooled to 0°C and ethyl ether (60ml) was added with vigorous stirring for 10 min. The organic phase was separated, washed twice with water then dried and the solvent evaporated under reduced pressure. The crude 4-acetyloxy eptanoic acid was dissolved in MeOH (78ml) and treated with 1N NaOH solution (39ml). The reaction mixture was concentrated until 1/3 volume and 2N HCl was added until pH 1. The reaction mixture was stirred for 1 h and extracted with Et $_{20}$, dried and filtered. The product was purified by bulb to bulb distillation under vacuo to give lactone 21 in 60% yield. ee 92% (glc analyses: Megadex column $T_{initial}=110^{\circ}C/3min / 3^{\circ}C/min/ T_{final}=180^{\circ}C)$.

<u>Correlation of lactones 14, 15 and 16</u>. The correlation of lactones 14, 15 and 16 were carried out in the same way as described for γ -lactone 13: 14 was transformed into the δ -lactone 23, 15 and 16 into the δ -lactones 35 and 36 respectively.

Intermediate product characterizations:

<u>Diol 24a</u>. ¹H NMR (CDCl₃, 90MHz) δ : 1.4-1.9 (8H, m), 2.6-2.9 (2H, m), 3.5-3.7 (3H, m), 7.1-7.3 (5H, m). [α]_D²⁵=+9.3^{*} (c 1.00, CHCl₃).

<u>Acetyl derivative 25a</u>. ¹H NMR (CDCl₃/D₂O, 90MHz) δ : 1.4-1.9 (8H, m), 2.0 (3H, s), 2.6-2.9 (2H, m), 3.5-3.7 (1H, m), 4.0-4.2 (2H, m), 7.2-7.3 (5H, m). $[\alpha]_D^{25}$ =+8.6° (c 1.00, CHCl₃).

 $\begin{array}{c} \underline{\text{Alcohol 26a.}} & {}^{1}\text{H NMR} \ (\text{CDCl}_{3}/D_{2}\text{O}, \ 90\text{MHz}) \ \delta: \ 0.9 \ (3\text{H}, \ t, \ J=6.0\text{Hz}), \ 1.2-1.9 \ (8\text{H}, \ m), \\ 2.5-2.9 \ (2\text{H}, \ m), \ 3.5-3.7 \ (1\text{H}, \ m), \ 7.1-7.3 \ (5\text{H}, \ m). \ [\alpha]_{D}^{25} = +14.0^{\circ} \ (c \ 1.00, \ \text{CHCl}_{3}). \\ \underline{\text{Acetyl derivative 27a.}} & {}^{1}\text{H NMR} \ (\text{CDCl}_{3}, \ 90\text{MHz}) \ \delta: 0.9 \ (3\text{H}, \ t, \ J=6.0\text{Hz}), \ 1.2-2.0 \ (8\text{H}, \ m), \\ \underline{\text{Acetyl derivative 27a.}} & {}^{1}\text{H NMR} \ (\text{CDCl}_{3}, \ 90\text{MHz}) \ \delta: 0.9 \ (3\text{H}, \ t, \ J=6.0\text{Hz}), \ 1.2-2.0 \ (8\text{H}, \ m), \\ \underline{\text{Acetyl derivative 27a.}} & {}^{1}\text{H NMR} \ (\text{CDCl}_{3}, \ 90\text{MHz}) \ \delta: 0.9 \ (3\text{H}, \ t, \ J=6.0\text{Hz}), \ 1.2-2.0 \ (8\text{H}, \ m), \\ \underline{\text{Acetyl derivative 27a.}} & {}^{1}\text{H NMR} \ (\text{CDCl}_{3}, \ 90\text{MHz}) \ \delta: 0.9 \ (3\text{H}, \ t, \ J=6.0\text{Hz}), \ 1.2-2.0 \ (8\text{H}, \ m), \\ \underline{\text{Acetyl derivative 27a.}} & \underline{\text{Acetyl derivative 27a.}} & \underline{\text{Acetyl derivative 27a.}} \ (8\text{H}, \ m), \ 1.2-2.0 \ (8\text{H}, \ m), \ 1.2-2-2.0 \ (8\text{H}, \ m), \ 1.2-2-2.$

<u>ACCUT derivative 27a</u>. H NMR (CDCl₃, 90MHz) $\delta:0.9$ (3H, t, J=6.0Hz), 1.2-2.0 (8H, m), 2.1 (3H, m), 2.4-2.7 (2H, m), 5.0 (1H, q, J=6.0Hz), 7.1-7.3 (5H, m). $[\alpha]_D^{25}$ =-13.2 (c 1.00, CHCl₃).

 $\begin{array}{c} \underline{\text{Diol 24b}} & {}^{1}\text{H NMR} \ (\text{CDECl}_{3}/D_{2}\text{O}, \ 250\text{MHz}) \ \delta: \ 1.40-1.80 \ (8\text{H}, \ \text{m}), \ 2.60 \ (2\text{H}, \ \text{t}, \ \text{J=7.50Hz}), \\ 3.50-3.68 \ (3\text{H}, \ \text{m}), \ 7.10-7.30 \ (5\text{H}, \ \text{m}). \ \left[\alpha\right]_{D}^{2\,5} = -1.4^{\circ} \ (\text{c} \ 1.00, \ \text{CHCl}_{3}). \\ \underline{\text{Acetyl derivative 25b.}} & {}^{1}\text{H NMR} \ (\text{CDCl}_{3}/D_{2}\text{O}, \ 250\text{MHz}) \ \delta: \ 1.35-1.85 \ (8\text{H}, \ \text{m}), \ 2.03 \ (3\text{H}, \ \text{m}). \ 1.35-1.85 \ (8\text{H}, \ \text{m}), \ 2.03 \ (3\text{H}, \ \text{m}). \ 1.35-1.85 \ (8\text{H}, \ \text{m}), \ 2.03 \ (3\text{H}, \ \text{m}). \ 1.35-1.85 \ (8\text{H}, \ \text{m}), \ 2.03 \ (3\text{H}, \ \text{m}). \ 1.35-1.85 \ (8\text{H}, \ \text{m}), \ 2.03 \ (3\text{H}, \ \text{m}). \ 1.35-1.85 \ (8\text{H}, \ \text{m}), \ 2.03 \ (3\text{H}, \ \text{m}). \ 1.35-1.85 \ (8\text{H}, \ \text{m}), \ 2.03 \ (3\text{H}, \ \text{m}). \ 1.35-1.85 \ (8\text{H}, \ \text{m}). \ 1.$

<u>ACCUVI GETIVELIVE 255</u>. "H NMR (CDC1₃/D₂O, 250MHz) &: 1.35-1.85 (8H, m), 2.03 (3H, s), 2.63 (2H, t, J=7.50Hz), 3.57-3.68 (1H, m), 4.07 (2H, t, J=6.50Hz), 7.14-7.31 (5H, m). $[\alpha]_D^{25}=-1.5^\circ$ (c 1.00, CHCl₃).

<u>Alcohol 26b</u>. ¹H MNR (CDCl₃/D₂O, 250MHz) δ : 0.91 (3H, t, J=7.25Hz), 1.30-1.85(8H, m), 2.63 (2H, t, J=7.50Hz), 3.57-3.68 (1H, m), 7.15-7.32 (5H, m). $[\alpha]_D^{25}=0^{\circ}$ (c 1.00, CHCl₃).

<u>Acetyl derivative 27b.</u> ¹H NMR (CDCl₃, 250MHz) δ : 0.88 (3H, t, J=7.00Hz), 1.20-1.70 (8H, m), 2.02 (3H, B), 2.58-2.64 (2H, m), 4.88-4.97 (1H, m), 7.13-7.32 (5H, m). $[\alpha]_D^{25}$ =-4.7° (c 1.00, CHCl₃).

<u>Diol 24c</u>. ¹H NMR (CDCl₃/D₂O, 250MHz) δ : 1.33-1.85 (10H, m), 2.61 (2H, t, J=7.50Hz), 3.50-3.62 (3H, m), 7.10-7.30 (5H, m). $[\alpha]_D^{2^5}=-2.1^\circ$ (c 1.00, CHCl₃).

<u>Acetyl derivative 25c</u>. ¹H NMR (CDCl₃/D₂O, 250MHz) &: 1.35-1.85 (10H, m), 2.03 (3H, s), 2.63 (2H, t, J=7.50Hz), 3.55-3.64 (1H, m), 4.05 (2H, t, J=6.50Hz), 7.11-7.29 (5H, m). $[\alpha]_D^{25}=-0.3^{\circ}$ (c 1.00, CHCl₃).

<u>Alcohol 26c</u>. ¹H NMR (CDCl₃/D₂O, 300MHz) δ : 0.90 (3H, t, J=7.50Hz), 1.25-1.85 (10H, m), 2.63 (2H, t, J=5.00Hz), 3.56-3.62 (1H, m), 7.13-7.29 (5H, m). $[\alpha]_D^{25}$ =+1.3° (c 1.00, CHCl₃).

 $\frac{\text{Acetyl derivative 27c.}}{(4H, m), 1.45-1.70 (6H, m), 2.01 (3H, s), 2.55-2.64 (2H, m), 4.85-4.95 (1H, m), 7.12-7.40 (5H, m). [<math>\alpha$] $_{D}^{25}$ =-4.4° (c 1.00, CHCl₃).

Lactol 29. A solution of lactone 13 (4.00g, 0.021mmol) in dry methylene chloride (200ml) at -78°C and under nitrogen was treated with a solution of DIBAL (31.6ml, 1M in n-hexane). After stirring for 1 h, the reaction mixture was quenched with 10% NaOH aqueous solution and extracted with methylene chloride. The organic extracts were washed with water, brine, dried over sodium sulfate and filtered. The solvent was evaporated under reduced pressure and the product was purified by flash chromatography (n-hexane/AcOEt 7/3). The lactol 29 was obtained, as diastereoisomeric mixture, in 94% yield. ¹H NMR (CDCl3/D₂O, 250MHz) δ : 1 62-2.12 (6H, m), 2.60-2.80 (2H, m), 3.95-4.30 (1H, m), 5.48-5.60 (1H, m), 7.13-7.32 (5H, m).

<u>Unsaturated alcohol 30</u>. Propyltriphenylphosphonium bromide (8.67g, 22.5mmol) dissolved in dry THF (220ml) under nitrogen was treated with a solution of n-BuLi (13.2ml, 1.5M in n-hexane) After stirring for 2 h at room temperature, the reaction mixture was cooled at -78°C and a solution of lactol **29** (1.73g, 19.0mmol) in dry THF (5ml) was added. The temperature was raised to room temperature. After stirring for 30 min at this temperature, the reaction mixture was quenched with a saturated NH4Cl solution and extracted with AcOEt. The organic extracts were dried, filtered and the solvent evaporated under reduced pressure. The crude product was purified by flash chromatography (n-hexane/AcOEt 8/2) to give the product **30** in 89% yield (Z/E mixture). ¹H NMR (CDC1₃/D₂O, 250MHz) δ : 0.98 (3H, t, J=7.50Hz), 1.46-1.62 (2H, m), 1.71.1.82 (2H, m), 1.93-2.23 (4H, m), 2.60-2.88 (2H, m), 3.60-3.70 (1H, m), 5.28-5.56 (2H, m), 7.15-7.35 (5H, m)

<u>Acetate 32</u>. Unsaturated alcohol 30 (1.20g, 5.45mmol) in 200 ml of $MeOH/H_{2O}$ (3:1) was hydrogenated in the presence of catalytic amounts of 10% Pd/C. After 20 h, the reaction

mixture was filtered and the solvent evaporated under reduced pressure to give the crude alcohol 31. 31: ¹H NMR (CDCl₃/D₂O, 90MHz) δ : 0.9 (3H, t, J=8.0Hz), 1.2-1.9 (12H, m), 2.6-2.8 (2H, m), 3.5-3.7 (1H, m), 7.1-7.3 (5H, m). A solution of the crude alcohol 31 in dry methylene chloride (32ml) was treated with Et₃N (1.69ml, 12.0mmol), Ac₂O (1.03ml, 10.9mmol) and a catalytic amount of DMAP. After stirring at room temperature for 1 h, the reaction mixture was washed with water, 5% aqueous HCl and water. The organic layer was dried over sodium sulfate , filtered and the solvent evaporated under reduced pressure. The crude product was purified by flash chromatography (n-hexame/AcOEt 95/5) to give the acetate 32 in 95% yield. ¹H NMR (CDCl₃, 250MHz) δ : 0.87 (3H, t, J=6.75Hz), 1.20-1.30 (10H, m), 1.50-1.61 (2H, m), 1.80-1.92 (2H, m), 2.04 (3H, s), 2.52-2.72 (2H, m), 4.88-4.98 (1H, m), 7.13-7.32 (5H, m). [α]D²⁵=-10.4* (c 1.07, CHCl₃). Anal.Calcd.for C_{17H26}O₂: C,77.82; H,10.01. Found: C,77.87; H,10.03.

<u> γ -Lactone</u> 28. Lactone 28 was obtained from 32 as already reported for the transformation of compound 20 into 21. The crude product was purified by Kugerlohr distillation (135-140 °C/15mmHg) to give the γ -lactone 30 in 65% overall yield. ¹H NMR (CDCl₃, 250MHz) &: 0.88 (3H, t, J=7.50Hz), 1.23-1.94 (11H, m), 2.25-2.40 (1H, m), 2.49-2.58 (2H, m), 4.43-4.55 (1H, m). $[\alpha]_D^{25}$ =-36.9° (c 1.03, MeOH); $[\alpha]_D^{25}$ =-33.8° (c 1.05, CHCl₃). The 97% ee was determined by glc analysis on chiral column as reported for lactone 21. Anal.Calcdfor C₁₀H₁₈O₂: C,70.55; H,10.68. Found: C,70.53; H,10.63.

<u>Benzylic bromination-dehydrobromination. General Procedure.</u> A solution of saturated lactone (1.58mmol) in CCl_4 (8ml) was treated with NBS (2.37mmol) and a catalytic amount of PhCO₃H. The reaction mixture was refluxed for 1 h, then filtered and the solvent evaporated under reduced pressure. The crude bromo derivative was dissolved in CHCl₃ (15.8ml) and treated with DBU (1.9mmol). After refluxing for 3 h, the reaction mixture was cooled and treated with 2N HCl until pH 5. The organic layer was washed with water and brine, then dried over sodium sulfate and the solvent evaporated under reduced presure. The product was purified by chromatography n-hexane/AcOEt 8/2.

presure. The product was purified by chromatography n-hexane/AcOEt 8/2. <u>Lactone 5</u>. 84% yield. $[\alpha]_D^{2^5}=+2.9^{\circ}$ (1.04, CHCl₃). 95%ee. Anal.Calcd.for C₁₂H₁₂O₂: C,76.57; H,6.44. Found: C,76.55; H,6.41.

<u>Lactone 6</u>. 30% yield. $[\alpha]_D^{25} = +0.6^{\circ}$ (c 1.01, CHCl₃). 99% ee. Anal.Calcd.for C₁₃H₁₄O₂: C,77.20; H,6.99. Found: C,77.23; H,7.02.

<u>Lactone 35</u>. 80% yield. ¹H MNR (CDCl₃, 250MHz) δ : 1.88-2.03 (1H, m), 2.25-2.40 (1H, m), 2.48-2.71 (4H, m), 4.58-4.68 (1H, m), 6.19 (1H, dt, J=15.75, 7.00Hz), 6.50 (1H, dt, J=15.75, 1.35Hz), 7.15-7.40 (5H, m). Anal.Calcd.for $C_{13}H_{14}O_2$: C,77.20; H,6.99. Found: C,77.25; H,6.95.

<u>Lactone 36</u>. 82% yield. [']H NMR (CDCl₃, 250MHz) δ : 1.50-1.68 (1H, m), 1.74-2.02 (3H, m), 2.40-2.70 (4H, m), 4.35-4.46 (1H, m), 6.23 (1H, dt, J=15.75, 7.00Hz), 6.49 (1H, d, J=15.75Hz), 7.19-7.39 (5H, m). $[\alpha]_D^{25}=-0.5^{\circ}$ (c 0.84, CHCl₃). Anal.Calcd.for C₁₄H₁₆O₂: C,77.75; H,7.46. Found: C,77.70; H,7.44.

<u>Tetrahydrofuran derivative 20</u>. A solution of diol 17 (500mg, 2.58mmol) in methylene chloride (26ml) was treated with Ph₃P (1.49g, 5.52mmol) and NBS (1.01g, 5.52mmol). After stirring for 48 h at room temperature the solvent was evaporated under reduced pressure. The crude mixture was taken up with Et₂O, filtered and the solvent evaporated under reduced pressure. The product was purified by flash chromatography (n-hexane-AcOEt 9/1) to give 22 in 70% yield. ¹H NMR (CDCl₃, 250MHz) &: 1.40-1.54 (1H, m), 1.69-2.05 (5H, m), 2.59-2.63 (2H, m), 3.68-3.94 (3H, m), 7.13-7.38 (5H, m). $[\alpha]_2^{25}=+4.2^{\circ}$ (c 1.06, CHCl₃). Anal.Calcd.for C₁₂H₁₆O: C,81.77; H,9.17. Found: C,81.74, H,9.17.

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